

**This Page Is Inserted by IFW Operations  
and is not a part of the Official Record**

## **BEST AVAILABLE IMAGES**

**Defective images within this document are accurate representations of the original documents submitted by the applicant.**

**Defects in the images may include (but are not limited to):**

- **BLACK BORDERS**
- **TEXT CUT OFF AT TOP, BOTTOM OR SIDES**
- **FADED TEXT**
- **ILLEGIBLE TEXT**
- **SKEWED/SLANTED IMAGES**
- **COLORED PHOTOS**
- **BLACK OR VERY BLACK AND WHITE DARK PHOTOS**
- **GRAY SCALE DOCUMENTS**

**IMAGES ARE BEST AVAILABLE COPY.**

**As rescanning documents *will not* correct images,  
please do not report the images to the  
Image Problem Mailbox.**

**THIS PAGE BLANK (USPTO)**

09/03853



INVESTOR IN PEOPLE



GB00/582

The Patent Office  
Concept House  
Cardiff Road  
Newport  
South Wales  
NP10 8QQ

## PRIORITY DOCUMENT

SUBMITTED OR TRANSMITTED IN  
COMPLIANCE WITH RULE 17.1(a) OR (b)

REC'D 08 MAR 2000

WIPO PCT

I, the undersigned, being an officer duly authorised in accordance with Section 74(1) and (4) of the Deregulation & Contracting Out Act 1994, to sign and issue certificates on behalf of the Comptroller-General, hereby certify that annexed hereto is a true copy of the documents as originally filed in connection with the patent application identified therein.

I also certify that the attached copy of the request for grant of a Patent (Form 1/77) bears an amendment, effected by this office, following a request by the applicant and agreed to by the Comptroller-General.

In accordance with the Patents (Companies Re-registration) Rules 1982, if a company named in this certificate and any accompanying documents has re-registered under the Companies Act 1980 with the same name as that with which it was registered immediately before re-registration save for the substitution as, or inclusion as, the last part of the name of the words "public limited company" or their equivalents in Welsh, references to the name of the company in this certificate and any accompanying documents shall be treated as references to the name with which it is so re-registered.

In accordance with the rules, the words "public limited company" may be replaced by p.l.c., plc, P.L.C. or PLC.

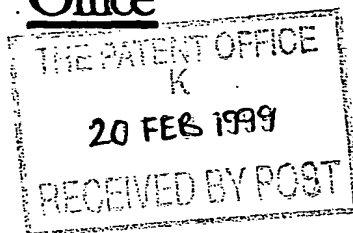
Re-registration under the Companies Act does not constitute a new legal entity but merely subjects the company to certain additional company law rules.

Signed

Dated 24 February 2000

**THIS PAGE BLANK (USPTO)**

The  
Patent  
Office



20 FEB 1999 E426788-2 D10007  
PO 000 0.00 - 9903805.1

1/77

# Request for grant of a patent

(See the notes on the back of this form. You can also get an explanatory leaflet from the Patent Office to help you fill in this form)

The Patent Office

Cardiff Road  
Newport  
Gwent NP9 1RH

1. Your reference

P1366

2. Patent application number

(The Patent Office will fill in this part)

9903805.1

3. Full name, address and postcode of the or of each applicant (underline all surnames)

THE UNIVERSITY OF SHEFFIELD  
WESTERN BANK  
SHEFFIELD  
S10 2TN

Patents ADP number (if you know it)

If the applicant is a corporate body, give the country/state of its incorporation

89/242002

4. Title of the invention

PLURIPOTENTIAL CELLS-1

5. Name of your agent (if you have one)

MARKGRAAF PATENTS LIMITED

Form S1/77 14/10/9

"Address for service" in the United Kingdom to which all correspondence should be sent (including the postcode)

THE CRESCENT  
54 BLOSSOM STREET  
YORK YO24 1AP  
UNITED KINGDOM

Harrison Goldard Foster  
Belmont House  
20 Wood Lane  
LEEDS  
LS6 2AE

Patents ADP number (if you know it)

2253003

7296486002

6. If you are declaring priority from one or more earlier patent applications, give the country and the date of filing of the or of each of these earlier applications and (if you know it) the or each application number

Country

Priority application number  
(if you know it)

Date of filing  
(day / month / year)

7. If this application is divided or otherwise derived from an earlier UK application, give the number and the filing date of the earlier application

Number of earlier application

Date of filing  
(day / month / year)

8. Is a statement of inventorship and of right to grant of a patent required in support of this request? (Answer 'Yes' if:

YES

- a) any applicant named in part 3 is not an inventor, or
  - b) there is an inventor who is not named as an applicant, or
  - c) any named applicant is a corporate body.
- See note (d))

700010 5-8578527  
**Patents Form 1/77**

9. Enter the number of sheets for any of the following items you are filing with this form.  
 Do not count copies of the same document

Continuation sheets of this form

Description

44

Claim(s)

Abstract

Drawing(s)

616

10. If you are also filing any of the following, state how many against each item.

Priority documents

Translations of priority documents

Statement of inventorship and right to grant of a patent (Patents Form 7/77)

Request for preliminary examination and search (Patents Form 9/77)

Request for substantive examination (Patents Form 10/77)

Any other documents (Please specify)

11.

I/We request the grant of a patent on the basis of this application.

Signature *Mark Graaf* Date 9.02.1999

MARKGRAAF PATENTS LIMITED

12. Name and daytime telephone number of person to contact in the United Kingdom

DR ROB C DOCHERTY

01904 610586

**Warning**

After an application for a patent has been filed, the Comptroller of the Patent Office will consider whether publication or communication of the invention should be prohibited or restricted under Section 22 of the Patents Act 1977. You will be informed if it is necessary to prohibit or restrict your invention in this way. Furthermore, if you live in the United Kingdom, Section 23 of the Patents Act 1977 stops you from applying for a patent abroad without first getting written permission from the Patent Office unless an application has been filed at least 6 weeks beforehand in the United Kingdom for a patent for the same invention and either no direction prohibiting publication or communication has been given, or any such direction has been revoked.

**Notes**

- If you need help to fill in this form or you have any questions, please contact the Patent Office on 0645 300303.
- Write your answers in capital letters using black ink or you may type them.
- If there is not enough space for all the relevant details on any part of this form, please continue on a separate sheet of paper and write "see continuation sheet" in the relevant part(s). Any continuation sheet should be attached to this form.
- If you have answered 'Yes' Patents Form 7/77 will need to be filed.
- Once you have filled in the form you must remember to sign and date it.
- For details of the fee and ways to pay please contact the Patent Office.

**Patents Form 1/77**

**PLURIPOTENTIAL CELLS-1**

5 The invention herein described relates to isolated pluripotent cells, comprising at least part of the cytoplasm from a teratocarcinoma cell and a nucleus of a somatic cell; methods to prepare such cells; therapeutic compositions of said cells; and uses thereof.

10 Animal embryonic development is a highly regulated development process that combines cell proliferation and cell/tissue differentiation to produce an intact organism. The co-ordination of cell proliferation and differentiation is, and has been, the subject of intense research and the information derived from this has contributed to our understanding of cell function and disease. For example and not by way of limitation, regulation of gene expression, cell  
15 differentiation, oncology, teratology.

Mammalian embryonic development is remarkably conserved during the early stages. Post fertilisation the early embryo completes four rounds of cleavage to form a morula of 16 cells. These cells complete several more rounds of  
20 division and develop into a blastocyst in which the cells can be divided into two distinct regions; the inner cell mass, which will form the embryo, and the trophoblast, which will form extra embryonic tissue, (eg placenta).

Those cells that form part of the embryo up until the formation of the  
25 blastocyst are said to be totipotent (e.g. each cell has the developmental potential to form a complete embryo and all the cells required to support the growth and development of said embryo).

During the formation of the blastocyst, the cells that comprise the inner cell mass (ICM) are said to be pluripotent (ie each cell has the developmental potential to form a variety of tissues).

- 5 Embryonic stem cells may be principally derived from two embryonic sources. Pluripotent cells isolated from the inner cell mass are termed embryonic stem cells (ES cells). An alternate source of pluripotent cells is derived from primordial germ cells isolated from the mesenteries or genital ridges of days 8.5-12.5 *post coitum* embryos which would ultimately  
10 differentiate into germ cells. These pluripotent cells are referred to as embryonic germ cells (EG cells). Each of these types of pluripotent cell has the same developmental potential with respect to differentiation into alternate cell types.
- 15 It is important to note that an intact embryo cannot be produced from a single pluripotent cell (eg either an ES or EG cell). Therefore a pluripotent cell has an increased commitment to terminal differentiation when compared to a totipotent cell.
- 20 Until very recently *in vitro* culture of human ES cells was not possible. The first indication that conditions may be determined which could allow the establishment of human ES cells in culture is described in WO 96/22362. The application describes cell lines and growth conditions which allow the continuous proliferation of primate ES cells which exhibit a range of  
25 characteristics or markers which are associated with stem cells having pluripotent characteristics.

For example, and not by way of limitation, the expression of specific cell surface markers SSEA-3 (+), SSEA-4 (+), TRA-1-60 (+), TRA-1-81 (+) (  
30 Shevinsky *et al* 1982; Kannagi *et al* 1983; Andrews *et al* 1984a; Thomson *et*



*al* 1995) and alkaline phosphatase (+). In addition the established primate cell lines disclosed in WO 96/22362 have stable karyotypes and continue to proliferate in an undifferentiated state in continuous culture. The primate ES  
5 cell lines also retain the ability, throughout their continuous culture, to form tissues derived from all three embryonic germ layers (endoderm, mesoderm and ectoderm).

More recently Thomson *et al* (Science 282: 1145-1147, 1998) have published  
10 conditions in which human ES cells can be established in culture. The above characteristics shown by primate ES cells are also shown by the human ES cell lines. In addition the human cell lines show high levels of telomerase activity, a characteristic of cells which show the ability to divide continuously in culture.

15 An alternative source of pluripotential embryonic stem cells are those derived from primordial embryonal germ cells (EG cells) which are located in the mesenteries or genital ridges of embryos. WO 98/43679 describes the isolation of EG cells from the gonadal or genital ridges of human embryos. EG cells  
20 described in WO 98/43679 exhibit features in common with primate and human ES cells, (eg expression of cell surface markers, continuous proliferation in culture in an undifferentiated state, normal karyotype and the ability to differentiated into selected tissues under defined conditions).

25 It is evident that the use of *in vitro* cultures of pluripotential stem cells, especially human cells, has important ramifications for both basic research (eg as a model for studying gene expression and/or tissue differentiation) and in transplantation and/or replacement therapies for tissues which have been damaged either through injury or disease. The establishment of *in vitro*  
30 cultures of human ES and EG cells is a major step toward realising the full

potential of this technology; because of their pluripotent nature ES and EG cells may be capable of differentiating under controlled conditions into a variety of cell types and/or tissues and organs that could have a wide variety of applications. For example, and not by way of limitation, replacement of  
5 damaged and/or diseased coronary and/or major arteries; replacement of damaged and/or diseased organs ( eg as a result of kidney disease, (eg cirrhosis), diabetes, various autoimmune diseases); replacement of damaged neurones ( eg Alzhiemers disease, Parkinsons disease, spinal injuries) or cancer. It will also be apparent to one skilled in the art that diseases such as  
10 AIDS may benefit from from tissues derived from ES or EG cells. The depletion of T-cells through virus induced cell death is the major contributory factor to the immuno-compromised state of AIDS suffers.

However, there are practical and ethical difficulties associated with the use of  
15 material derived from human embryos. Moreover, such allogeneic material, if transplanted into another human, may illicit a severe immune reaction in the host and be thus destroyed.

It has been known for many years that amphibian somatic cell nuclei retain  
20 their ability to give rise to entire organisms when they are transplanted into egg cells which have had their nucleus removed or inactivated (Gurdon 1974). Thus determination of the pluripotent of these cells must be controlled by the egg cytoplasm which was able to in effect reprogramme the somatic cell nucleus into a totipotent state.

25 Mammalian somatic cell nuclei have also been shown to retain this placidity and can be reprogrammed when transferred to enucleated oocytes, (Campbell *et al*; Wakayama *et al*)

Moreover nucleated mouse ES cells have been shown to be able to reprogramme somatic cell nuclei, although in this case, a heterokaryon was produced containing the cytoplasm and nuclei from both types of cells so it is difficult to determine the actual mechanism of action of the reprogramming state.

In all these examples, although the material produced is genetically identical to the somatic cell donor, these somatic cells were reprogrammed by cellular elements are derived from either oocytes or ES cells and again, in human this poses practical and ethical concerns.

Embryonal carcinoma cells derived from teratomas are also able to reprogramme somatic cell nuclei in order to produce cells with pluroptential characteristics.

Teratomas, tumours that contain a wide range of more or less organised tissues have been known in humans for many hundreds of years. They typically occur as gonadal tumours of both men and women, but they also occur in other sites. The gonadal forms of these tumours are generally believed to originate from germ cells, and the extra-gonadal forms, which typically have the same range of histology, are widely thought to arise from 'mis-placed' germ cells that have migrated incorrectly during embryogenesis. However, a non-germ cell origin from persisting embryonic stem cells may be considered in some cases, especially for teratomas occurring in the new born.

Because of the presumption of a germ cell origin in most cases, teratomas are generally classed as germ cell tumours (GCT), which also manifest a range of other histological types, including seminoma (often called dysgerminoma in females), embryonal carcinoma (EC), yolk sac carcinoma and

choriocarcinoma. GCT may contain any combination of these tissue types, with or without elements of teratoma. Combinations of teratoma and EC are frequently described as teratocarcinoma. Commonly, human GCT are divided into pure seminomas, in which none of the other histological types occur, and  
5 non-seminomatous GCT (NSGCT), which may contain any combination of histological types, with or without elements of teratoma. Confusingly, NSGCT may also contain elements of seminoma.

Ovarian GCT are most commonly benign and contain only well differentiated  
10 somatic tissues that may include bone, muscle, nerve. Often well-organised tissues are found, including teeth and hair. By contrast, human testicular GCT are always malignant containing any combination of the tumour tissue types discussed above. Any teratoma elements present may be less well organised than in benign human ovarian teratomas, but somatic cell types such as nerve,  
15 muscle, bone and cartilage may be quite recognisable. Testicular GCT are rare but have a peak incidence after puberty, being the most common form of cancer in young men between the ages of 20 - 35.

Early histopathological studies of human GCT led to the proposal that EC  
20 cells resemble early embryonic cells and are the stem cells that give rise to all the other cell types in GCT, with the exception of seminoma. (In the currently prevailing view, seminoma more closely resembles the primordial germ cells from which these tumours arise, and so may represent an earlier stage in tumour development and progression). Detailed experimental study of GCT  
25 became possible with the discovery that male laboratory mice of the strain 129 develop spontaneous testicular teratocarcinomas. Studies of these mice confirmed that these tumours had a germ cell tumour origin, and indeed arose from primordial germ cells (PGC) at about the time that they migrated into the genital ridge of the developing embryo - 11 - 13 days of gestation in the

laboratory mouse. Evidence was also obtained supporting the hypothesis that EC cells are pluripotent stem cells that are able to generate the whole range of differentiated cells found in teratocarcinomas. Subsequently it was found that similar teratocarcinomas and teratomas could be derived from embryos that have been transplanted to ectopic sites.

For reasons that remain unclear, seminoma has not been observed in laboratory mice, and so no animal model of this tumour exists. Spermatocytic seminoma do occur in dogs but, although these also occur in old men, they appear to be a quite different tumour type with different aetiology to the GCT.

EC cell lines that can be maintained in vitro have been derived from several mouse teratocarcinomas. Some of these EC cell lines were found to retain a pluripotent phenotype and could be induced to differentiate into a range of cell types, either by transplantation back to a syngeneic mouse, in which case a teratocarcinoma would be formed, or by various manipulations in vitro. Some EC cell lines differentiated spontaneously when allowed to grow to confluence. Others required a feeder layer of irradiated or mitomycin-treated cells if they were to retain an EC phenotype, and they differentiated spontaneously if removed from the feeder cells. In a number of cases, it was found that maintaining EC cells in suspension culture for several days forced them to form floating clumps of cells that became vesiculated and began to differentiate. These floating clumps were known as embryoid bodies; a wide range of differentiated cell types, including nerve and muscle, would grow out from these if they were subsequently allowed to attach to a tissue culture surface. It was also found that a number of chemical agents, most strikingly retinoic acid, also induced the differentiation of many murine EC cells into a range of cell types. The precise cell types formed in response to any of these treatments depended upon the particular EC cell line.

Access to EC cell lines allowed detailed characterisation of their properties and pattern of marker expression. Several surface antigens, notably the 'F9 antigen' and Stage Specific Embryonic Antigen-1 (SSEA-1) were identified as characteristically expressed by mouse EC cells. It was further noted that these

5 cells did not express class 1 major histocompatibility (MHC) antigens - H-2, in the mouse. These features, as well as morphology and their capacity to differentiate suggested that murine EC cells resembled cells of the inner cell mass or primitive ectoderm of the early mouse embryo. This resemblance was confirmed by the finding that some EC cells could differentiate and participate

10 in normal embryonic development, when transferred to a blastocyst of an early mouse embryo, which was then re-implanted in a pseudo-pregnant mother and allowed to develop to term. In some cases the chimeras developing from such EC cell ↔ embryo combinations were normal with extensive contributions from the EC cell component; in some cases the chimeras subsequently

15 developed teratocarcinomas, indicating that the tumour phenotype of the EC cells had not been fully suppressed. Only in a very small number of cases was germ cell chimerism reported.

EC cells have been shown to have many of the features which characterise ES

20 and/or EG cells. EC cells are relatively easy to establish in culture, express cell surface markers associated with ES and/or EG cells, can be maintained in continuous culture in an undifferentiated state and have the potential to differentiate into selected tissues both *in vivo* and *in vitro*. However what is also evident is that EC cells contain a mutation, or mutations, in genes

25 (oncogenes) which result in the additional undesirable feature that the cells retain the potential to form tumours.

It has been known for several years that selected chemical treatments of cells in culture can result in cells extruding nuclei resulting in the formation of

separate nuclear and cytoplasmic parts termed karyoplasts and cytoplasts, respectfully. It is well known in the art that the separated parts of the cell may be reconstituted via cell fusion. For example, and not by way of limitation it is possible to produce a cytoplast from one cell and fuse the cytoplast to a selected cell to form a cytoplasmic hybrid, or as is commonly known, a cybrid. In addition it is also possible to fuse the karyoplast to a selected cell to form a nuclear hybrid. The nuclei fuse after nuclear membrane breakdown during mitosis and reconstitute after cytokinesis to form a polyploid or aneuploid nucleus. These techniques are well known in the art and will not be detailed extensively at this stage.

We have prepared cytoplasts derived from EC cells and fused the cytoplasts to form cybrids with selected somatic cells. The aim of this approach is to re-programme the differentiated somatic cell nucleus, through contact with factors located in the EC cytoplasm, so that, the cybrid de-differentiates and so takes on the characteristic features of a pluripotent cell. This then provides the basis for the establishment of pluripotent cell lines which, upon exposure to various differentiation factors, can lead to the production of selected differentiated tissue for use in, amongst other things, transplantation therapy.

Advantageously there is no requirement to use harvested embryonic cells to derive the cytoplasts. Therefore any ethical issues with respect to the use of embryos in this way is circumvented. In addition the establishment of EC cells in culture is a relatively amenable task when compared to the problems of establishing human stem cell cultures *in vitro*. Finally the removal of the EC cell nucleus from the donating EC cell results in no transfer of genetic material carrying potential oncogenes to the cybrid cell so formed.

It is therefore an object of the invention to provide a pluripotential cell that is not derived from embryonic tissue from a primary source.

5 It is a further object of the invention to provide methods of combining at least part of the cytoplasm of an EC cell with a somatic nucleus.

It is yet a further object of the invention to provide a pluripotential cell having the capacity to differentiate into selected tissues upon exposure to appropriate factors.

10

According to a first aspect of the invention there is provided a cell comprising at least part of the cytoplasm derived from at least one embryonal carcinoma cell combined with at least the nucleus of at least one somatic cell.

15 In a preferred embodiment of the invention said cell, ideally a cybrid, is characterised by the possession of at least one pluripotential characteristic.

We believe that the acquisition of this pluripotential characteristic is as a result of the re-programming of said somatic nucleus.

20

It will be apparent to those skilled in the art that the cell of the invention may be derived, most preferably, by the creation of a cybrid; but an alternative option involves the fusion of a somatic cell with an EC cell. Clearly this latter option is not preferred because of the potentially oncogenic genome (gene) of  
25 the donating EC cell. Preferably the further step of removing the EC nucleus is described hereinafter.

Ideally said pluripotential characteristic includes the ability to differentiate into at least one selected tissue type, preferably upon exposure to at least one  
30 differentiation factor.



Alternatively, or additionally, said pluripotential characteristic includes the ability of said cell to proliferate in culture in an undifferentiated state.

- 5 In yet a further preferred embodiment of the invention said cell has the capacity to proliferate in continuous culture in an undifferentiated state for at least 6 months and ideally 12 months.

- Alternatively or additionally said pluripotential characteristic includes the  
10 expression of at least one selected marker of pluripotential cells.

- In yet still a further preferred embodiment of the invention said selected marker is a cell surface marker. Preferably said cell surface marker is selected from the group including SSEA-1 (-); SSEA-3 (+); SSEA-4 (+); TRA-1-60  
15 (+); TRA-1-81 (+); alkaline phosphatase (+).

- Alternatively, or additionally, said pluripotential characteristic includes the presence of telomerase activity in said pluripotential cell. Ideally said telomerase activity is correlated with extension of telomeres.  
20

- For the sake of clarity, telomerase enzymes add, *de novo*, repetitive DNA sequences to the ends of chromosomes. These ends are referred to as telomeres. For example the telomeres of human chromosomes contain the sequence '5 TTAGGG 3' repeated approximately 1000 times at their ends. In  
25 young, dividing cells the telomeres are relatively long. In aging, or non-dividing cells, the telomeres become shortened and there is a strong correlation between telomere shortening and capacity to proliferate. Methods to increase the length of telomeres to increase proliferative capacity are known in the art and are described in WO9513383.

Alternatively, or additionally, said pluripotential characteristic includes the presence of a chromosomal methylation pattern characteristic of pluripotential cells.

- 5 It is well known in the art that the genome of eukaryotic organisms is variably methylated through the addition of methyl ( -CH<sub>3</sub>) groups attached to cytosine residues in DNA to form 5'methylcytosine ( 5'-mC). Methylation is correlated with the control of gene expression. Typically genes that are hypomethylated tend to be highly expressed. Hypermethylation is correlated with reduced gene  
10 expression. It will be apparent to one skilled in the art that pluripotential cells will have a typical methylation pattern. This pattern may be analysed at a genomic level or at the level of a specific gene. Methods to analyse the extent of methylation are well known in the art and include, by example and not by way of limitation, restriction enzyme digestion of DNA with methylation  
15 sensitive restriction endonucleases followed by Southern blotting and probing with suitable gene probes ( Umezawa *et al* 1997).

Alternatively or additionally said pluripotential characteristic includes the ability to induce tumours when introduced into an animal, ideally a rodent  
20 experimental model. More ideally still said animal is immunosuppressed.

According to a second aspect of the invention there is provided a cell-line comprising cells according to the invention. Ideally, said cell-line are of human origin.

25

According to a third aspect of the invention there is provided a method for preparing a cytoplasm, or part thereof, for use in the production of the cell or cell line of invention comprising;

- 30 i) providing at least one EC cell;

- ii) separating at least part of the cytoplasm from the nucleus of said EC cell;
- iii) isolating said cytoplasmic part; and, optionally
- iv) storing said isolated cytoplasmic part prior to use.

5

In a preferred method of the invention said cytoplasmic part is a cytoplast.

It will be apparent to one skilled in the art that said cytoplast may be provided either as an aliquot isolated from at least one EC cell ( eg an aliquot extracted from an intact EC cell via micromanipulation techniques) or alternatively, and preferably, said cytoplasmic part may be provided as an isolated cytoplast.

In a preferred method of the invention said cytoplast part is separated from said nucleus by exposure to a pharmacologically effective amount of cytochalasin B.

It is well known in the art that cytochalasin B is an example of a chemical effective at separating the nucleus of a cell from the cytoplasm to form a karyoplast and cytoplast respectively, (Methods in Enzymology Vol 151, p221-237 1987).

According to a fourth aspect of the invention there is provided a method for preparing a cell or cell-line in accordance with the invention comprising;

- i) combining at least one EC cell with at least one somatic cell;
- 25 ii) removing from said combined cell, the EC cell nucleus;
- iii) culturing said cell under conditions conducive to proliferation and expansion of said cell; and, optionally
- iv) storing said cell culture under suitable conditions.

It will be apparent to one skilled in the art that methods of micromanipulation exist that facilitate the removal of nuclei from selected cells. It will be apparent that this method of the invention advantageously provides that ;

- 5    i)      the factors produced by the EC cell are continually produced thereby maintaining a steady-state level of factors necessary to reprogramme the somatic cell nucleus; and
- ii)     the EC cell nucleus is removed from the combined cell prior to mitosis ensuring oncogenic genes are not transferred.

10

It will be apparent to those skilled in the art that the nature of the somatic cell selected is not critical to the operation of the invention although the cell-type will be selected so as to optimise or maximise success in terms of production of a cell or cell-line of the invention.

15

According to a fifth aspect of the invention there is provided a method of combining at least part of the cytoplasm of an EC cell with a somatic cell comprising;

- i)      providing at least part of the cytoplasm of an EC cell;
- 20    ii)     combining said cytoplasmic part with at least one somatic cell;
- iii)    growing said combined cell in culture; and, optionally
- iv)     storing said combined cell under suitable storage conditions.

     In a preferred method of the invention said cytoplasmic part is provided as a  
25    cytoplasm.

     In yet a further preferred method of the invention said cytoplasm is combined with said somatic cell via cytoplasm/somatic cell fusion.

In the above described methods the EC cell and somatic cell are, ideally of human origin.

According to a sixth aspect of the invention there is provided a cell culture  
5 comprising at least one cell according to the invention.

According to a seventh aspect of the invention there is provided a method for inducing differentiation of at least one cell of the invention comprising:

- 10 i) providing a cell according to the invention;
- ii) culturing said cell under conditions conducive to the differentiation of said cell into at least one tissue; and, optionally
- iii) storage of said differentiated tissue prior to use under suitable storage conditions.

15

Ideally said culture conditions are selected so as to provide a tissue type, by example and not by way of limitation, that is, neuronal, muscle (eg smooth, striated, cardiac), bone, cartilage, liver, kidney, respiratory epithelium, haematopoietic cells, spleen, skin, stomach, intestine.

20

According to a eighth aspect of the invention there is provided at least one tissue type or organ comprising at least one cell according to the invention.

It will be apparent to one skilled in the art that differentiated tissue according  
25 to the invention may have extensive application with respect to transplantation therapy. For example, and not by way of limitation, replacement of damaged and/or diseased coronary and/or major arteries; replacement of damaged and/or diseased organs ( eg as a result of kidney disease, (eg cirrhosis), diabetes, various autoimmune diseases); replacement of damaged neurones ( 30 eg Alzhiemers disease, Parkinsons disease, spinal injuries) or cancer. It will

also be apparent to one skilled in the art that diseases such as AIDS may benefit from tissues derived from the cells of the invention. The depletion of T-cells through virus induced cell death is the major contributory factor to the immuno-compromised state of AIDS suffers. The provision of a non-exhaustive supply of T-cells derived from a non-infected somatic cell from the patient has obvious benefits. Moreover, tissue rejection due to host cell immune responses are likely to be negligible since the somatic nucleus used in the cybrid would ideally be derived from the patient requiring the replacement tissue or organ.

10

According to an ninth aspect of the invention there is provided a therapeutic composition comprising at least one cell of the invention including a suitable excipient, diluant or carrier.

15 In a preferred embodiment of the invention said therapeutic composition is provided for use in tissue transplantation.

According to a tenth aspect of the invention there is provided a method to treat conditions or diseases requiring transplantation of tissue comprising;

20

- i) providing at least one tissue type or organ according to the invention;
- ii) surgically introducing said tissue type or organ to a patient to be treated; and
- iii) treating said patient under conditions which are conducive to the acceptance of said transplanted tissue by said patient.

25

According to a eleventh aspect of the invention there is provided a kit comprising; at least one cell according to the invention; instructions with respect to the maintenance of said cell in culture; and, optionally, factors

required to induce differentiation of said cell to at least one desired tissue type or organ.

An embodiment of the invention will now be described by example only and  
5 with reference to the following tables and figures wherein;

Table 1 represents a summary of the human EC cell lines and some of the characteristic features of said human EC cell-lines ;

10 Table 2 represents a summary of the murine EC cell lines used;

Figure 1 shows various human teratocarcinoma-derived cell lines. The characteristic embryonal carcinoma (EC) morphology of several human EC cell lines derived from testicular (a-e) and extragonadal (f-g) teratocarcinomas: (a) 1156QE, (b) TERA-1, (c) SuSa, (d) 833KE, (e)  
15 1777NRpmet, (f) 1618K, (g) NCCIT. Bar = 50 $\mu$ m;

Figure 2 shows a flow cytofluorimetric analyses of surface antigen expression by the human EC cell line 2102Ep;

20

Figure 3 shows differentiation of several human EC cell lines caused by alteration in growth conditions: (a) 2102Ep cells placed at 10<sup>5</sup> per 75 cm<sup>2</sup> flask (low density); (b) spontaneous differentiation of a few cells in a culture of 1156QE; (c) a culture of SuSa cells passaged by trypsinisation; (d) a culture of  
25 1777NRp differentiated cells derived from 1777NRpmet EC cells by repeated passage of low density (Bronson et al 1983a). Bar =50 $\mu$ m; and

Figure 4 shows differentiation of TERA-2 derived human EC cells: (a) NTERA-2 cl D1 human EC cells; (b) parental TERA-2 culture, showing  
30 mixed patches of EC and non-EC cells; (c) neurons and other differentiated

cells arising in cultures of NTERA-2 cl D1 cells following induction with retinoic acid (Andrews 1984); (d) non-neural differentiated NTERA-2 cl D1 cells induced by exposure to hexamethylene bisacetamide. (Bar = 50 $\mu$ m);

- 5 Figure 5 shows heterokaryons obtained by fusion of 2102Ep cl 4D3 human EC cells and the human T-cell leukaemia cell line, MOLT4. Several heterokaryons containing 2 and 3 nuclei can be seen in this field. (Bar = 50 $\mu$ m); and

- 10 Figure 6 shows cytoplasts derived from NTERA-2 cl D1 human EC cells following enucleation with cytochalasin B. A remaining nucleated cell (top right) has been included in the field for comparison.

### **Materials and Methods**

#### **Enucleation of cells to yield 'cytoplasts' and 'karyoplasts' or 'mini-cells'.**

- 15 One of the techniques that is employed in our method for producing Re-Programmed Embryonic Stem Cells ( RPES cells) is the use of cytochalasin B to generate enucleated EC cells (EC cytoplasts) as the cytoplasm donor, and 'karyoplasts' (also called 'mini-cells') from the differentiated or committed cells as the nucleus donor. Cytochalasin B is well-known to induce cells to
- 20 extrude their nuclei (Carter, 1967) and has been employed by numerous authors to induce enucleation of a wide range of cells of a variety of species including both mouse and human cells (Poste 1972; Prescott et al 1972; Goldman et al 1973; Wright and Hayflick 1973; Ege and Ringertz 1974a; Wigler and Weinstein 1975). Such enucleation results in a cell lacking a
- 25 nucleus, but is otherwise intact and viable for a number of days (Goldman et al 1973); these enucleated cells have been called anucleate cells (Poste 1972) or cytoplasts (Veomett et al 1974). The nucleus that is extruded from the cell retains a thin rim of cytoplasm and is surrounded by a plasma membrane;



these structures have been called 'karyoplasts' (Veomett et al 1974) or 'mini-cells' (Ege and Ringertz 1975). Enucleation of cells to yield both cytoplasts and karyoplasts may be achieved by well-established techniques in which cells growing attached to a plastic disc are inverted over a solution of cytochalasin B in a centrifuge tube and centrifuged; the cytoplasts remain attached to the plastic disc, while the karyoplasts are pelleted at the bottom of the centrifuge tube (Prescott et al 1972). Alternatively, cells in suspension may be centrifuged through a density gradient, typically composed of Ficoll, containing cytochalasin B (Wigler and Weinstein 1975). In this case, cytoplasts and karyoplasts are formed and may be recovered from different parts of the gradient after centrifugation.

Using the method described by Prescott et al 1972 NTERA-2 c1D1 EC cells growing on a plastic disc were inverted over a solution of 7.5 $\mu$ g/ml cytochalasin B, in phosphate buffered saline containing 10% fetal bovine serum, in a 50 ml centrifuge tube, and centrifuged for 30 minutes at 12,000 rpm and 37°C in a J2 Bectman Centrifuge using a JA20 rotor. Cytoplasts without nuclei remain attached to the disc. Occasional cells that have escaped enucleation also remain, please see Figure 6. After recovery by incubation overnight in medium without cytochalasin, the discs were fixed in methanol and stained with haematoxylin and cosin (Bar - 50 $\mu$ m).

#### Methods for combining (fusing) the cytoplasm of one cell with the nucleus of another.

The methods for creating hybrid cells by fusing two or more cells of different origins together are very well established and widely known. For a review of

the commonly used methods based upon Sendai virus induced cell fusion, or cell fusion induced by polyethylene glycol (PEG), see Kennett (1979).

Briefly, mixtures of cells that it is desired to fuse are incubated with a fusogenic agent, such as Sendai virus or PEG, often with centrifugation or agitation to encourage clumping and close apposition of the cell membranes; variables such as time, temperature, cell concentration and fusogenic agent concentration are optimised for each cell combination. An example of cell fusion to produce heterokaryons is presented in Figure 6. Cell fusion was carried out using Polyethylene Glycol 1000, as described by Kennett 1979. Following fusion, the cells were seeded into tissue culture dishes, and incubated in fresh medium overnight. They were then fixed with methanol and stained with haematoxylin and eosin. Several heterokaryons containing 2 and 3 nuclei can be seen in this field. (Bar = 50 $\mu$ m).

These techniques have also been shown to allow fusion of cytoplasts, prepared by cytochalasin B induced enucleation, with whole cells or karyoplasts, also derived by cytochalasin B induced enucleation (Poste and Reeve 1971; Ege and Ringertz 1975; Ege et al 1973, 1974; Veomett et al 1974; Wright and Hayflick 1975; Shay 1977)).

20

Another technique that is now well established and widely used for inducing cell fusion, 'electrofusion', involves passing short electric pulses through mixtures of cells (Neil and Zimmermann 1993).

25

### **Production of RPES cells**

The production of RPES cells requires several steps:

1. the selection of appropriate differentiated cells (the Nucleus Donor) and, if necessary, the isolation of their nuclei,
2. the selection of EC cells (the Cytoplasm Donor),
3. the fusion of the differentiated cell nuclei with the EC cells, and
- 5        4. the removal of the EC cell nucleus, either before or after fusion.

The production technique may, in some cases, be optimised by pre-treatment of the differentiated cells, or contemporaneous treatment of the differentiated cell/ EC cell fused products, with various agents such as, but not limited to, inhibitors of DNA methylation, to enhance the ability of the differentiated cell  
10        nucleus to be re-programmed. After the production of the RPES cells additional methods are required to propagate the cells, to characterise their properties and to induce them to differentiate into required somatic cell types.

#### 15        **Differentiated cells to be used as Nuclear Donors**

A large range of somatic cells derived from any tissue or organ of an adult mammal or human, or from embryos or foetuses, or from extra-embryonic tissues such as the trophoblast or yolk sac may be used as a source of nuclei for reprogramming. Particular somatic cell types include but are not limited to  
20        thymocytes, peripheral blood lymphocytes, epidermal cells such as from the bucal cavity, cumulus cells, or other stem cells isolated from biopsies of various tissues, such as the bone marrow, the nervous system and the gut. The technique may also be applied to various established cell lines, such as those derived from various tumours including, for example, but not limited to  
25        lymphoblastoid cell lines. The selected somatic cells used for the re-programming procedure may be used directly upon isolation or they may be cultured for a short time before further manipulation. In some instances such somatic cells may be combined entirely with EC cells as described below, or

nuclei or karyoplasts may first be isolated from them, for example using agents such as cytochalasin B, as discussed above, or by other methods. For example, nuclei may also be isolated using established micromanipulation procedures, or other established cell fractionation procedures.

5

### **Parental EC cells to be used as Cytoplasm Donors**

A large number of EC cell lines have been isolated from human teratocarcinomas, which occur predominantly, but not exclusively, as testicular germ cell tumours. Examples of available human EC cell lines are shown in Table 1. Similarly, EC cell lines derived from teratocarcinomas of the laboratory mouse have also been derived and are readily available. Examples of available mouse EC cell lines are shown in Table 2. To date EC cell lines have not been described from any other species, but if they were derived in the future we would anticipate that they should behave in a manner similar to the existing human and mouse EC cells which resemble one another. Thus, newly isolated EC cells of other species, or from human or mouse sources, should also be able to re-program differentiated cells to RPES cells as described in its proposal for human and mouse EC cells.

Human EC cells can be readily recognised by a combination of features that include their morphology (see Figure 1), their expression of the cell surface antigens SSEA3 (Shevinsky et al 1982, Andrews et al 1982, 1984b, 1996), SSEA4 (Kannagi et al 1983, Andrews et al 1984b, 1996), and TRA-1-60 and TRA-1-81 (Andrews et al 1984a, 1984b, 1996), and typically by low expression or absence of SSEA1 (Andrews et al 1980, 1982, 1984b, 1996) (see Figure 2). By contrast, mouse EC cells typically express SSEA1 (Solter and Knowles, 1978) but not SSEA3 (Shevinsky et al 1982), SSEA4 (Kannagi et al 1983) or TRA-1-60 or TRA-1-81 (Andrews et al 1984a). Human EC

cells like mouse EC cells express high levels of alkaline phosphatase (ALP) (Bernstine et al 1973, Benham et al 1981); in the case of human EC cells most ALP activity is due to expression of the liver/bone/kidney isoform which can be detected as a cell surface antigen by monoclonal antibodies TRA-2-59 and TRA-2-54 (Andrews et al 1984c). In common with ES cells and primordial germ cells, EC cells also typically express the transcription factor Oct3/4 (Rosfjord and Rizzino 1994; Brehm et al 1998).

#### Fusion of parental differentiated cells and parental EC cells to yield RPES cells

Several methods may be used to combine the cytoplasm of an EC cell and the nucleus of a differentiated cell to yield an RPES containing the nuclear genome of the differentiated cell but not the EC cell.

A. Cells may be fused by use of chemical agents such as polyethylene glycol (PEG) or viruses such as Sendai virus, or by passing an electric current through a mixture of cells. As discussed above, these methods are well known and may be readily applied. These methods may be used to fuse:

1. a differentiated cell with an EC cell, or
2. a karyoplast from a differentiated cell with an EC cell, or
3. a differentiated cell with one or more cytoplasts isolated from EC cells, or
4. a karyoplast from a differentiated cell with one or more cytoplasts isolated from EC cells.

In cases (1) and (2), the result will initially be a heterokaryon containing two nuclei, one from each parental cell. If this heterokaryon

were allowed to divide the result would be a hybrid cell containing a single nucleus with a complete or partial genome from each parental cell. However, in our method of producing RPES cells, the EC nucleus is removed prior to cell division of the hybrid cell, so that the derivative dividing cell population retains only the genome of the parental differentiated cell.

In cases (3) and (4) the EC nucleus is removed from the EC cell before fusion, for example by enucleation with cytochalasin B as discussed above, so that the resulting product contains only the differentiated cell nucleus and cytoplasm from the EC cell parent. In any of these cases, the resulting RPES cells that continue to proliferate retain only the nuclear genome of the differentiated parental cell, which is now reprogrammed to express a new pattern of gene activity.

In cases (1) and (2) the EC cell nucleus is removed from the heterokaryon in one of several ways that include, but are not limited to, partial enucleation using drugs such as cytochalasin B, applied in the same manner as described above for enucleating EC cells and generating cytoplasts for fusion. In the present case in which enucleation is carried out after fusion, some heterokaryons lose both nuclei, in which case they do not proliferate, some heterokaryons lose the differentiated cell nucleus, in which case they retain the parental EC nucleus and continue proliferating, some heterokaryons lose the EC cell nucleus, in which case they continue proliferating as RPES cells, and some heterokaryons retain both nuclei and eventually continue proliferating as hybrid cells. Several methods are used to select the RPES cells and to eliminate any of the cells retaining an EC cell genome or to eliminate any cells retaining a somatic nucleus that has

failed to undergo re-programming. In one method, the proliferating cells are cloned by established techniques (e.g. by picking single cells with a micropipette - see Andrews et al 1982, 1984b), and individual clones are screened using genetic markers for those that retain an EC genome. The latter cells are discarded, whereas those that retain only a differentiated cell genome but not an EC cell derived genome, and express an RPES phenotype, are retained. Standard DNA genotyping techniques using well established DNA fingerprinting technology (Jeffreys et al 1985, 1988; Yan et al 1996) may be used to identify whether the nuclear genome of any proliferating cells is derived from either the EC cell or differentiated cell parent, or both.

In another method, before use as a fusion partner, the EC cell parent is genetically marked by insertion of a gene that will allow selection against any cell carrying that gene; for example, the EC cell can be stably transfected with a vector encoding the Herpes Simplex Virus-1 Tk gene (HSV1-Tk), such that any cells carrying that gene can be killed by culture in the presence of a number of drugs including acyclovir (9-[(2-hydroxyethoxy)methyl]guanine) or FIAU (1-(2-deoxy-2-fluoro- $\beta$ -D-arabinofuranosyl)-5-iodouracil) (Borrelli et al 1988; Hasty et al 1991), or gancyclovir (Rubinstein et al 1993; McCarrick and Andrews 1992). In this method, following partial enucleation, the remaining heterokaryons are cultured in medium containing this drug, and only those that have lost the EC cell nucleus survive. Other selectable genetic systems can also be similarly used. Persisting parental differentiated cells that have not been reprogrammed are removed by cloning the surviving cells, or by selecting RPES cells by virtue of their expression of specific surface antigen markers that include, but are not limited to, SSEA3, SSEA4, TRA-1-60 or TRA-1-81, as discussed

above as characteristic markers of EC cells. These same markers have been shown to be expressed by ES cells derived directly from human embryos (Thomson et al 1998; Shambloott et al 1998) For the latter approach, fluorescence activated cell sorting (FACS), a widely used method for separating subsets of cells can be used (e.g. Andrews et al 1982, 1987; Ackerman et al 1994; Williams et al 1988).

In another method, the EC cell parent is incubated prior to fusion, with a drug that irreversibly inactivates its nucleus and prevents its replication, for example, topoisomerase inhibitors such as etoposide (Downes et al 1991; Fulka and Moor 1993). The resulting heterokaryon naturally eliminates this treated nucleus prior to cell division, so that the resulting dividing cell population only contains the genome derived from the parental differentiated cell. This approach may also be combined with the preceding 'partial enucleation of heterokaryons' approach to ensure complete loss of the EC genome.

In another method, after cell fusion to produce a heterokaryon, the EC cell nucleus is removed by micro-manipulation.

B. Rather than chemical, viral or electrically induced fusion, the nucleus of the differentiated cell is combined with an EC cell parent by micro-manipulation. In this method, the nucleus of the differentiated cell is withdrawn using a micropipette inserted through the cell membrane. It is then injected either into an inoculated EC cell, or into an intact EC. In the later case the EC cell nucleus is then removed by a similar technique, or by one of the techniques described above, before nuclear fusion and cell division occurs.



### **Growth and selection of RPES cells**

Following fusion to combine a differentiated cell and an EC cells, with prior or subsequent removal of the EC cell nucleus, it is necessary to provide  
 5 appropriate conditions for the re-programming of the differentiated cell nucleus and for the subsequent proliferation of the resulting RPES cells.

Several methods are used to enhance the efficiency of reprogramming:

- 10 1. prior to fusion the differentiated cell and EC cell are synchronised with respect to position in the cell cycle, by use of reversible inhibitors that arrest the cell cycle at specific stages (e.g. nocodazole), or by the use of conditions such as low serum to arrest cells in G1, or by selection of cells at specific stages of the cell cycle by using vital DNA stains and flow microfluorimetry  
 15 (Fluorescence Activated Cell Sorting) (Ashihara and Baserga 1979; Andrews et al 1987; Crissman 1995; Stein et al 1995).
- 20 2. the differentiated cell or the immediate fusion product is cultured in the presence of drugs that inhibit methylation or promote demethylation (e.g. 5-azacytidine) (e.g. Taylor and Jones 1979; Jones 1985; Keshet et al 1986), or alter the structure of chromatin, for example butyrate, spermine, trichostatin A or trapoxin which inhibit deacetylation and promote acetylation of histones, which plays a role in X chromosome inactivation, gene imprinting and regulation of gene expression (Caldarera et al 1975; McKnight et al  
 25 1980; Stein et al 1997; Hu et al 1998; Wolffe and Pruss 1996;).
3. the period of time between production of heterokaryons and the removal of the EC cell nucleus is made as long as possible without permitting nuclear fusion. This period can be elongated by

culturing the heterokaryons under conditions that reversibly inhibit progress through the cell cycle (e.g. thymidine block - Stein et al 1995), or by altering growth conditions, such as serum starvation or lowered temperature, that retard cell division but permit reprogramming to proceed.

4. any, or all combinations of these methods.

In all these experiments the cells are cultured in standard cell culture media that include but are not restricted to Dulbecco's modified Eagle's Medium (DME, high glucose formulation) or Ham's F12, supplemented in some cases with foetal bovine serum or with other additives (e.g. see Andrews et al 1980, 1982, 1984, 1994). Subsequent to fusion and re-programming, the growth of the resulting cells may be optimised culture on feeder layers of cells that include, but are not restricted to, irradiated or mitomycin C treated STO cells, or embryonic fibroblasts of various species, including humans (see Robertson 1987a; Thomson et al 1998). The cells may be cultured in the presence of various growth factors or other tissue culture additives, that include but are not restricted to LIF, FGF, SCF

## 20 Differentiation of the RPES cells

In the best cases, the RPES cells acquire pluripotent properties that closely resemble those of embryonic stem cells, so that the RPES cells are able to differentiate and to initiate differentiation pathways that result in the formation of any cell type that may be found in the adult, embryo or in extra-embryonic tissues, given appropriate conditions. The maintenance of an EC cell state can be monitored by assay of various markers that include the cell surface antigens SSEA3, SSEA4, TRA-1-60, TRA-1-81, by their expression of alkaline phosphatase and by expression of Oct3/4, as discussed above. The RPES cells

typically retain their stem cell phenotype when cultured on appropriate feeder cells. However, they can initiate differentiation under a variety of circumstances.

- 5 Thus removal from feeder cells, or culture in suspension, followed by replating in the absence of feeder cells in appropriate tissue culture flasks results in differentiation of stem cells into a variety of cell types that include neurons, muscle of various sorts and haematopoietic cells (see descriptions in Robertson 1987a). Differentiation of pluripotent stem cells (e.g. see Figures 3  
10 and 4) may also be initiated by altered conditions affecting cell density and aggregation (e.g. seeding at low cell densities or trypsinisation) or exposure to various agents that include but are not restricted to retinoic acid, and other retinoids, hexamethylene bisacetamide, and the bone morphogenetic proteins (see Robertson 1987a; Andrews 1984; Andrews et al 1982, 1990, 1994, 1996;  
15 Thomson et al 1998). The type of cells that arise depend upon the nature of the inducing agent, and the culture conditions including the presence or absence of specific growth factors or other molecules.

### **Discussion**

- 20 Although pluripotent stem cell lines have been derived from early embryos (Robertson, 1987b; Thomson et al 1995, 1998), primordial germ cells (Matsui et al 1992; Shamblott et al 1998) and from germ cell tumours (reviewed, Andrews, 1998) of various species, including the laboratory mouse, rhesus monkeys and humans, and nuclei from differentiated somatic adult cells have  
25 been re-programmed to yield embryonic stem cells by transplantation to enucleated oocytes (Campbell et al 1996; Wakayama et al 1998), there are no reports that pluripotent stem cells, resembling embryonic stem cells with the capacity to differentiate into a variety of functional somatic cell types, can be

produced by the re-programming of differentiated or committed embryonic or adult somatic cells, or extra-embryonic cells, without the use of oocytes.

We now describe methods by which embryonal carcinoma (EC) cells, derived from teratocarcinomas, can be used to re-program various somatic, differentiated cells, or other embryonic or extra-embryonic cell types, to a state from which they can then be induced to differentiate into one or more functional differentiated cell types that are distinct from the parental cells. In the best cases, but not necessarily in all cases, the re-programmed cells produced by this technique, called 'Re-programmed Embryonic Stem Cells' (RPES cells), resemble embryonic stem cells derived directly from early embryos, and can be induced to differentiate into a broad range of functional, differentiated cell types that include, but are not limited to, neurons, muscle (including skeletal and cardiac muscle) and haematopoietic cells. These RPES cells are diploid with a normal karyotype, and isogenic with the differentiated parental cells from which they are derived. They may be used to generate differentiated cells for transplantation and use in cell and tissue replacement therapies.

In some cases, only partial reprogramming occurs with, for example, the activation of several genes that are not active in the parental differentiated nuclear donor cell. Such cells are also of use in a variety of these same circumstances.

In our method, RPES cells are created by combining the nucleus from a differentiated or committed cell (the Nuclear donor), whether from adults or from embryos, with the cytoplasm from an EC cell (the Cytoplasm donor), from which the nucleus is removed. Several methods can be used to combine the nucleus from the differentiated cell and the cytoplasm from the EC cell; in some methods the EC cell nucleus is removed prior to combination of the cytoplasm with the donated nucleus, and in other methods the EC cell nucleus

is removed after combination. If EC cells and differentiated cells from the same species are used, then the resulting RPES cells retain cytoplasmic genetic determinants (e.g. the mitochondrial genome) and a nuclear genome from the same species. By contrast, embryonic stem-like cells produced by

5 transplantation of somatic cells into enucleated oocytes of other species will continue to harbour mitochondria of that other species. Especially for the production of human RPES cells and their differentiated derivatives for transplantation into a human host, the maintenance of a human nuclear and human cytoplasmic genome could be a distinct advantage. Further, RPES

10 cells that are isogenic with the anticipated human host can be produced by this technique without resort to any embryo, so avoiding practical difficulties that may be associated, for example, with immune rejection upon transplantation to the human host, and also obviating ethical difficulties inherent in the use of human embryos.

15

The method that we describe incorporates the techniques for maintaining and propagating the RPES cells produced, and the techniques for inducing them to differentiate into a range of differentiated, functional cell types.

## 20 **References**

- Andrews P.W. and Goodfellow P.N. (1980) Antigen expression by somatic cell hybrids of a murine embryonal carcinoma cell with thymocytes and L cells. *Somatic Cell Genet.* **6**: 271-284.
- 25 Andrews P.W., Bronson D.L., Benham F., Strickland S. and Knowles B.B. (1980) A comparative study of eight cell lines derived from human testicular teratocarcinoma. *Int. J. Cancer* **26**: 269-280.

Andrews P.W., Goodfellow P.N., Shevinsky L., Bronson D. L. and Knowles B.B. (1982) Cell surface antigens of a clonal human embryonal carcinoma cell line: Morphological and antigenic differentiation in culture. *Int. J. Cancer* 29: 523-531.

- 5 Andrews P.W. (1982) Human embryonal carcinoma cells in culture do not synthesize fibronectin until they differentiate. *Int. J. Cancer* 30: 567-571.

Andrews P.W., Damjanov I., Simon D., Banting G., Carlin C., Dracopoli N.C. and Fogh J. (1984b) Pluripotent embryonal carcinoma clones derived from the human teratocarcinoma cell line Tera-2: Differentiation in vivo and in vitro. *Lab. Invest.* 50: 147-162.

- 10

Andrews P.W., Meyer L.J., Bednarz K.L. and Harris H. (1984c) Two monoclonal antibodies recognizing determinants on human embryonal carcinoma cells react specifically with the liver isozyme of human alkaline phosphatase. *Hybridoma* 3: 33-39.

- 15 Andrews P.W. (1984) Retinoic acid induces neuronal differentiation of a cloned human embryonal carcinoma cell line in vitro. *Dev. Biol.* 103: 285-293.

Andrews P.W., Banting G.S., Damjanov I., Arnaud D. and Avner P. (1984a) Three monoclonal antibodies defining distinct differentiation antigens associated with different high molecular weight polypeptides on the surface of human embryonal carcinoma cells. *Hybridoma* 3: 347-361.

- 20

Andrews P.W., Gönczöl E., Plotkin S.A., Dignazio M. and Oosterhuis J.W. (1986) Differentiation of TERA-2 human embryonal carcinoma cells into neurons and HCMV permissive cells: Induction by agents other than retinoic acid. *Differentiation* 31: 119-126.

- 25

Andrews, P.W., Oosterhuis J.W. and Damjanov I. (1987) Cell lines from human germ cell lines. In: *Teratocarcinomas and embryonic stem cells: A practical approach* (E.J. Robertson, ed.). IRL Press, Oxford, pp 207-248.

- Andrews P.A., Nudelman E., Hakomori S. -i. and Fenderson B.A. (1990). Different patterns of glycolipid antigens are expressed following differentiation of TERA-2 human embryonal carcinoma cells induced by retinoic acid, hexamethylene bisacetamide (HMBA) or bromodeoxyuridine (BUdR). Differentiation 43, 131-138.
- Andrews P.W., Damjanov I., Berends J., Kumpf S., Zappavigna V. Mavilio F. and Sampath K. (1994). Inhibition of proliferation and induction of differentiation of pluripotent human embryonal carcinoma cells by osteogenic protein-1 (or bone morphogenetic protein-7). Laboratory Investigation, 71, 243-251.
- Ashihara, J., and Baserga, R. (1979). Cell Synchronisation Methods. Enzymology, 58, 248-262.
- Bernstine, E.F., Hooper, M.L., Grandchamp, S., Ephrussi, B. (1973). Alkaline phosphatase activity in mouse teratoma. Proc Natl Acad Sci USA 70, 3899-3902.
- Brehm, A., Ovitt, C.E., Schöler, H.R. (1978). Oct 4: more than just a powerful marker of the mammalian germline? Acta Path. Microbiol, Immunol, Scand. 106, 114-126.
- Bronson, D.L., Andrews, P.W., Solter, D., Cervenka, J., Lange, P.H., and Fraley, E.E. (1980). A cell line derived from a metastasis of a human testicular germ-cell tumour. Cancer Res. 40, 2500-2506.
- Bronson, D.L., Ritzi, D.M., Fraley, E.E., and Dalton, A.J. (1978). Morphologic evidence for retrovirus production by epithelial cells derived from a human testicular tumour metastasis. J. Natl. Cancer Inst. 60, 1305-1308.
- Bronson, D.L., Andrews, P.W., Vessella, R.L., and Fraley, E.E. (1983). In vitro differentiation of human embryonal carcinoma cells. In

"Teratocarcinoma Stem Cells" (L.M. Silver, G.R. Martin and S. Strickland, eds.), Cold Spring Harbor Conferences on Cell Proliferation, Vol. 10, pp. 597-605. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.

- 5 Bronson, D.L., Saxinger, W.C., Ritzi, D.M., Fraley, E.E. (1984). Production of virions with retrovirus morphology by human embryonal carcinoma cells in vitro. *J. Gen. Virol* 65, 1043-1051.

- 10 Campbell, K.H.S., McWhir, J., Ritchie, W.A., Wilmut, I. (1996). Sheep cloned by nuclear transfer from a cultured cell line. *Nature*, 380, 64-66.

Carter, S.B. (1967). Effects of cytocholasins on mammalian cells. *Nature*, 213, 261-264.

- 15 Downes, C.S., Mullinger, A.M., Johnson, R.T. (1991). Inhibitions of DNA topoisomerase II prevent sister chromatid separation in mammalian cells but do not prevent exit from mitosis. *Proc. Nat. Acad. Sci (USA)*. 88, 8895-8899.

- 20 Ege, T., and Ringertz, N.R., (1974). Preparation of microcells by enucleation of micronucleate cells. *Exp. Cells Res.* 87, 378-382.

Ege, T., Krondahl, U., Ringertz, N.R. (1974). Introduction of nuclei and micronuclei into cells and enucleated cytoplasms by Sendai virus induced fusion. *Exp Cell Res.* 88, 428-432.

25

Ege, T., and Ringertz, N.R. (1975). Viability of cells reconstituted by virus-induced fusion of mini cells with enucleate cells. *Expl. Cell Res.*, 94, 469-473.

- 30 Ege, T., Zeuthen, J., Ringertz, N.R. (1973). Cell fusion with enucleated cytoplasms. *Nobel*, 23, 189-194.



- Engström, W., Tally, M., Granerus, M., Hedley, E.P., and Schofield, P. (1991). Growth factors and the control of human teratoma cell proliferation. *Recent Res. Cancer Res.* 123, 145-153.
- 5 Fenderson, B.A., Andrews, P.W., Nudelman, E., Clausen, H., and Hakomori, S.-I., (1987). Glycolipid core structure switching from clobo- to lacto- and ganglio-series during retinoic acid-induced differentiation of TERA-2 derived human embryonal carcinoma cells. *Dev. Biol.* 122, 21-34
- 10 Fogh, J., and Trempe, G. (1975). New human tumor cell lines. In "Human Tumor Cells in vitro" (J.Fogh, ed.), pp. 115-159. Plenum Press, New York.
- Fulka, J., Moor, R.M. (1993). Non invasive chemical enucleation of mouse  
15 oocytes. *Mol. Reprod. Devel.*, 34, 427-430.
- Goldman, R.D., Plllack, R., Hopkins, N.H. (1973). Presrvation of normal behaviour by enucleated cells in culture. *Proc. Nat. Acad. Sci. (USA)*. 70, 750-754.
- 20 Gönczöl, E., Andrews, P.W., and Plotkin, S.A. (1984). Cytomegalovirus replicates in differentiated but not undifferentiated human embryonal carcinoma cells. *Science* 224, 159-161.
- 25 Gurdon J. The control of gene expression in animal development (Oxford University Press 1974).
- Hogan, B., Fellous, M., Avner, P., and Jacob, F. (1977). Isolation of a human teratoma cell line which expresses F9 antigen. *Nature (London)* 270, 515-518.

Jakob, H., Boon, T., Galliard, J., Nicolas, J.F., Jacob, F. (1973).  
Tératocarinome de la souris: isolement, culture et propriétés de cellules à  
potentialités multiples. *Ann Microbiol (Paris)* 124b 269-282.

- 5    Jeffreys, A.J., Wilson, V., Thein, S.L. (1985). Individual specific fingerprints  
of human DNA. *Nature*, 316, 76-79

Jeffreys, A.J., Wilson, V., Newmann, R., Keyte, J. (1988). Amplification of  
human minisatellites by the polymerase chain reaction - towards DNA  
10    fingerprinting of single cells.

Kennett, R.H. (1979). Cell Fusion in: *Cell Culture, Methods in Enzymology*.  
(eds. Jakoby, W.B., and Pastan, I.H.) Academic Press San Diego, 58, 345-359.

- 15    Matsui, Y., Szebo, K., and Hogan, B.L.M. (1992). Derivation of pluripotential  
embryonic stem cells for murine primordial germ cells in culture. *Cell*, 70,  
841-847.

Mavilio F., Simeone A., Boncinelli E. and Andrews P.W. (1988) Activation  
20    of four homeobox gene clusters in human embryonal carcinoma cells induced  
to differentiate by retinoic acid. *Differentiation*, 37,73-79.

Miller., W.H., and Dmitrovsky, E. (1991). Growth factors in human germ cell  
cancer. *Recent Res.Cancer Res.* 123, 183-189.

25

Neil, G.A., and Simmermann, U. (1993). Electrofusion, *Methods in*  
*Enzymology*, 220, 174-196.

Poste, G., and Reeve, P. (1971). Formation of hybrid cells and heterokaryons by fusion of enucleated and nucleated cells. *Nature New Biology*, 229, 122-125.

- 5 Poste, G. (1972). Enucleation of mammalian cells by cytochalasin B. *Exp. Cell Res.* 73, 273-286.

Prescott, D.M., Myerson, D., Wallace, J. (1972). Enucleation of mammalian cells with cytochalasin B. *exp. Cell Res.* 71, 480-485.

10

Robertson, E.J. (Editor) (1987). *Teratocarcinomas and embryonic stem cells: A practical approach*. IRL Press, Oxford.

- Robertson, E.J., (1987b). Embryo-derived stem cell lines in (Robertson, E.J.,  
15 Editor) *Teratocarcinomas and embryonic stem cells: A practical approach*. IRL Press, Oxford, pp 71-112.

- Rosfjord, E., and Rizzino A. (1994). The octamer motif present in the Rex-1 promoter binds Oct-1 and Oct-3 expressed by EC cells and ES cells. *Biochem.*  
20 *Biophys. Res. Counc.*, 203, 1795-1803.

- Shamblott, M.J., Axelman, J., Wang, S., Bugg, E.M., Littlefield, J.W.,  
Donovan, P.J., Blumenthal, P.D., Huggins, G.R., Gearhart, J.D. (1998).  
Derivation of pluripotent stem cells from cultured human primordial germ  
25 cells. *Proc. Nat. Acad. Sci. USA*, 95, 13726-13731.

Shay, J.W. (1997). Selection of recontributed cells from karyoplasts fused to chloramphenicol resistant cytoplasts. *Proc. Nat. Acad. Sci. USA*. 74, 2461-2464.

Shevinsky, L.H., Knowles, B.B., Damjanov, I., Solter, D. (1982). Monoclonal antibody to murine embryo defines a stage specific embryonic antigen expression on mouse embryos and human teratocarcinoma cells. *Cell* 30, 697-705.

5

Simeone, A., Acampora, D., Arcioni, L., Andrews, P.W., Boncinelli, E., and Mavilio, F. (1990). Sequential activation of human HOX2 homeobox genes by retinoic acid in human embryonal carcinoma cells. *Nature (London)* 346, 763-766.

10

Solter, D., and Knowles, B.B. (1978). Monoclonal antibody defining a stage specific mouse embryonic antigen (SSEA-1). *Proc Natl Acad Sci (USA)*. 75, 5565-5569.

15 Teshima, S., Shimosato, Y., Hirohashi, S., Tome, Y., Hayashi, I., Kanazawa, H., and Kakizoe, T., (1988). Four new human germ cell tumor cell lines. *Lab. Invest.* 59, 328-336.

Thomson, J.A., Itskovitz-Eldor, J., Shapira, S.S., Waknitz, K.A., Swiergiel, J.J., Marshall, V.S., Jones, J.M. (1998). Embryonic stem cell lines derived from human blastocysts. *Science*, 282, 1145-1147.

20 Thompson, S., Stern, P.L., Webb, M., Walsh, F.S., Engström, W., Evans, E.P., Shi, W.K., Hopkins, B., and Graham, F.F. (1984). Cloned human teratoma cells differentiate into neuron-like cells and other cell types in retinoic acid. *J. Cell Sci.* 72, 37-64.

25 Veomett, G., Prescott, D.M., Shay, J., Porter, K.R. (1974). Reconstruction of mammalian cells from nuclear and cytoplasmic components separated by treatment with cytocholasin B. *Proc Nat Acad Sci*, 71, 1999-2002.

30

- Vogelzang, N., Andrews, P.W., and Bronson, D. (1983). An extragonadal human embryonal carcinoma cell line 1618K. *Proc. Am. Assoc. Cancer Res.* 24, 3.
- 5 Von Keitz, A.T., Riedmiller, H., Neuman, K., Grutschart, W., Fonatsch, C. (1994). Establishment and characterisation of a seminoma cell line (S2). *Invest. Urol (Berlin)*. 5, 28-31.
- 10 Wakayama, T., Perry, A.C.F., Succotti, M., Johnson, K.R., Yanagimachi, R. (1998). Full term development of mice from enucleated oocytes injected with cumulus cell nuclei. *Nature*, 394, 369-374.
- 15 Wakeman, J.A., Heath, P.R., Pearson, R.C.A., Andrews, P.W. (1997) MAL mRNA is induced during the differentiation of human embryonal carcinoma cells into neurons, and is also localised within specific regions of the human brain. *Differentiation* 62:97-105.
- 20 Wakeman, J.A., Walsh, J., Andrews, P.W., (1998). Human Wnt-13 is developmentally regulated during the differentiation of NTERA-2 pluripotent human embryonal carcinoma cells. *Oncogene* 17:179-186
- Wang, N., Trend, B., Bronson, D.L., and Fraley, E.E. (1980). Nonrandom abnormalities in chromosome 1 in human testicular cancers. *Cancer Res.* 40, 796-802.
- 25 Wang, N., Perkins, K.L., Bronson, D.L., and Fraley, E.E. (1981). Cytogenetic evidence for pre-meiotic transformation of human testicular cancers. *Cancer Res.* 41, 2135-2140.

Wenk, J., Andrews, P.W., Casper, J., Hata, J-I., Pera, M.F., von Keitz, A., Damjanov, I., Fenderson, B.A. 1994. Glycolipids of germ cell tumours: extended globo-series glycolipids are a hallmark of human embryonal carcinoma cells. *Int. J. Cancer.*, 58, 108-115.

5

Wigler, M.H., and Weinstein, I.B. (1975). A preparative method for obtaining enucleated mammalian cells. *Biochem Biophys Res Comm.* 63, 669-674.

Williams B.P., Daniels G.L., Pym B., Sheer D., Povey S., Okubo Y., Andrews  
10 P.W. and Goodfellow P.N. (1988) Biochemical and genetic analysis of the OKa blood group antigen. *Immunogenetics*, 27, 322-329.

Wright, W.E., Hayflick, L. (1973). Enucleation of cultured human cells. *Proc. Soc. Exp. Biol. and Med.* 144, 587-592.

15

Wright, W.E., and Hayflick L. (1975). Use of biochemical lesions for selection of human cells with hybrid cytoplasms. *Proc. Nat. Acad. Sci (USA)*. 72, 1812-1816.

20 Yan, R., Ottenbreit, K., Hukku, B., Mally, M., Chou, S.R., Kaplan, J. (1996). DNA fingerprinting of human cell lines using PCR amplification of fragment length polymorphisms. *In Vitro Celular and Developmental Biology*, 32, 656-662.

25

**Table 1**  
**Human EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)**

Cell Line	Phenotype in Culture <sup>1,2</sup>	Surface Antigen Expression <sup>3,5</sup>					Tumour Origin		Xenograft	References
		SSEA3	SSEA4	TRA-1-60	TRA-1-81	TRA-2-49 TRA-2-54 (L-ALP) <sup>4</sup>	Original Site	Biopsy Site		
TERA2 including sublines such as NTERA-2 and clones	EC	+	+	+	+	+	Testis	Lung Metastasis	EC,T	Fogh & Trempe 1975 Wang <i>et al</i> 1981 Andrews 1984 Andrews <i>et al</i> 1984b, 1990 Gönczl <i>et al</i> 1984 Thompson <i>et al</i> 1984 Fenderson <i>et al</i> 1987 Simeone <i>et al</i> 1990 Engstrom <i>et al</i> 1991 Miller & Dmitrovsky 1991 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
2102Ep	EC	+	+	+	+	+	Testis	Primary	EC	Andrews <i>et al</i> 1980, 1982 Wang <i>et al</i> 1980,1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
2102E Rpmet	EC	+	+	+	+	+	Testis	Retroperitoneal lymphnode	EC,T	Wang <i>et al</i> 1980,1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
1156QE	EC	+	+	+	+	+	Testis	Primary	EC,C,S	Andrews <i>et al</i> 1980 Wang <i>et al</i> 1980,1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994

Table 1

Human EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)

Cell Line	Phenotype in Culture <sup>1,2</sup>	Surface Antigen Expression <sup>3,5</sup>					Tumour Origin			Xenograft	References
		SSEA3	SSEA4	TRA-1-60	TRA-1-81	TRA-2-49 TRA-2-54 (L-ALP) <sup>4</sup>	Original Site	Biopsy Site	Histology		
1218E	EC	+	+	+	+	+	Testis	Primary	EC,S	EC	Andrews <i>et al</i> 1980 Wang <i>et al</i> 1980, 1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
833KE	EC	+	+	+	+	+	Testis	Primary	EC,T,C,S	EC	Bronson <i>et al</i> 1978, 1980, 1984 Andrews <i>et al</i> 1980, 1996 Wang <i>et al</i> 1980, 1981 Wenk <i>et al</i> 1994
TERA1	EC	+	+	+	+	+	Testis	Lung metastasis	EC,T		Fogh & Trempe 1975 Wang <i>et al</i> 1981 Bronson <i>et al</i> 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
SuSa	EC	+	+	+	+	+	Testis		EC,T		Hogan <i>et al</i> 1977 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
NCC-IT	EC	+	+	+	+	+	Extragenodal	Primary	EC,T,Y	EC,T,C,Y	Teshima <i>et al</i> 1988 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
1618K	EC	+	+	+	+	+	Extragenodal		EL		Vogelzang <i>et al</i> 1983 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
1777N Rpmet	EC	+	+	+	+	+	Testis	Retroperitoneal Lymph node	EC		Bronson <i>et al</i> 1983, 1984 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994
S2	? not EC	-	-	+/-	-	-	Testis	Primary	Seminoma	?EC features	Von Keitz 1994 Wenk <i>et al</i> 1994 Andrews <i>et al</i> 1996 Wenk <i>et al</i> 1994



**Table 1**  
**Human EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)**

**Notes**

- 1 The phenotype of cells in culture is based upon observations of morphology, growth patterns and antigen expression (see, Andrews *et al* 1996).
- 2 Culture conditions: Human EC cells can generally be maintained in Dulbecco's Modified Eagles medium (DMEM), high glucose formulation, supplemented with glutamine and 10% foetal bovine serum, but other media have also been used (e.g. RPMI for NCCIT) (see Andrews and Damjanov 1994). High cell densities ( $75 \times 10^6$  per  $75 \text{ cm}^2$  tissue culture flask) are optimal for maintaining an EC phenotype (see Andrews *et al* 1982, 1984b, Andrews and Damjanov 1994). All the cell lines may be harvested for passage using 0.25% trypsin and 2mM EDTA, in  $\text{Ca}^{2+}/\text{Mg}^{2+}$  - free Dulbecco's phosphate buffered saline, but in some cases (TERA-2 and derivatives, and SuSa) clumping of cells is preferable for best maintenance of an EC phenotype. In the latter cases, cells are harvested for passage by scraping, for example with glass beads, rather than by use of trypsin (see Andrews *et al* 1984b, Andrews and Damjanov 1994).
- 3 Surface antigen expression: SSEA-3, SSEA-4, TRA-1-60 and TRA-1-81 expression is characteristic of human EC cell lines but the level of expression of these antigens is variable and appears to reflect their state of differentiation (e.g. see Andrews *et al* 1996, Andrews *et al* 1982, Kannagi *et al* 1983, Andrews *et al* 1984a, Shevinsky *et al* 1982).
- 4 High levels of the liver/bone/kidney isoforms of alkaline phosphatase (L-ALP) detected as a cell surface antigen by monoclonal antibodies TRA-2-49 and TRA-2-54 are also characteristic of human EC cells (Benham *et al* 1981, Andrews *et al* 1984c, Andrews *et al* 1996).
- 5 Differentiation and loss of EC phenotype: Many human EC cells undergo morphological changes and change in surface antigen expression, notably the induction of SSEA-1 and down regulation of SSEA-3, when cultured at low cell densities and well dispersed (Andrews *et al* 1982, 1984b). These changes appear to represent a limited capacity for differentiation. Other lines differentiate extensively if exposed to agents such as retinoic acid (e.g. NTERA-2, Andrews 1984; NCCIT, Teshima *et al* 1988), hexamethylene bisacetamide (e.g. NTERA-2, Andrews *et al* 1986, 1990), and the bone morphogenetic proteins (e.g. NTERA-2, Andrews *et al* 1994). Differentiation in the latter cases is marked by loss of the characteristic EC marker antigens, appearance of new antigens (e.g. see, Fenderson *et al* 1987), activation of new genes (e.g. *Hox* genes, Mavilo *et al* 1988; *Mal*, Wakeman *et al* 1997; *Wnt-13*, Wakeman *et al* 1998).

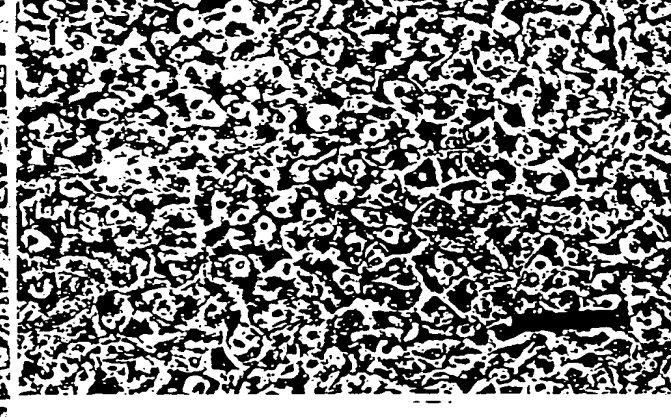
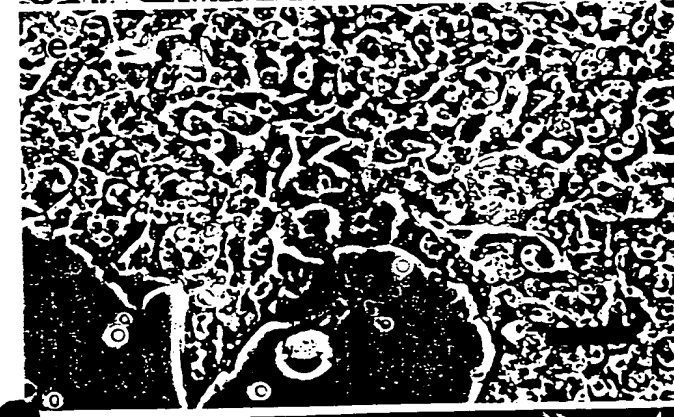
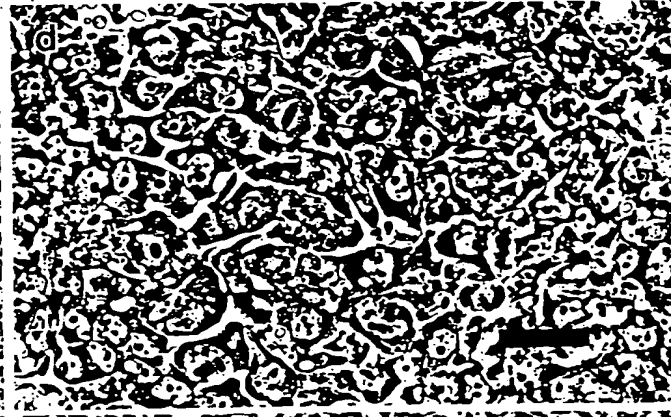
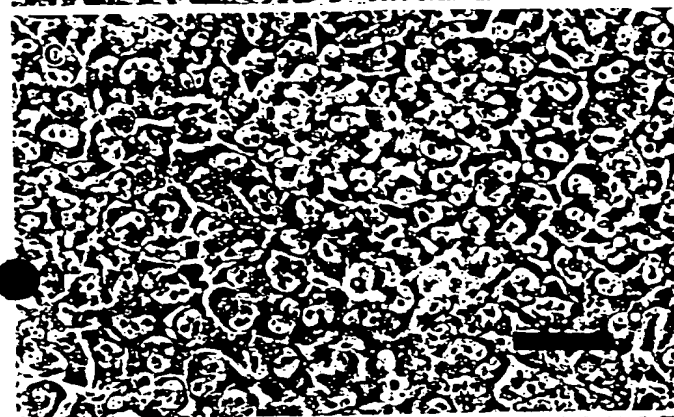
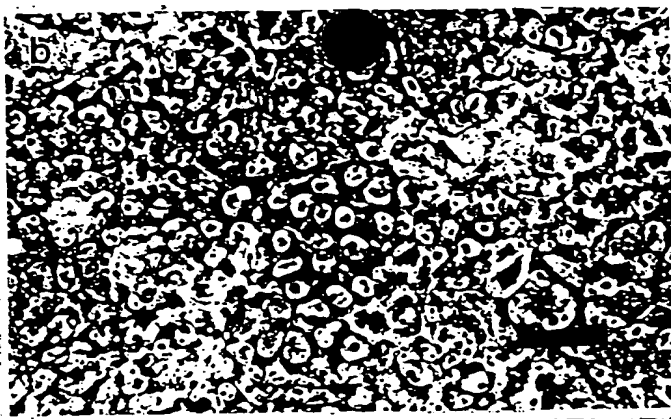
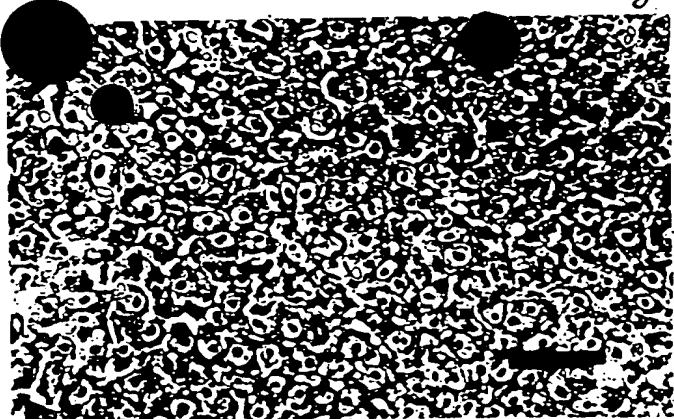
Table 2

Mouse EC Cell Cytoplasmic Donors (this list is illustrative but not comprehensive)

Cell Line	Surface Antigen Expression SSEA-1 <sup>1</sup>	Reference
F9	+	Bernstine <i>et al</i> 1993, Solter and Knowles 1978
PCC4	+	Jakob <i>et al</i> 1973, Solter and Knowles 1978
PCC3 (ND1)	+	Jakob <i>et al</i> 1973, Solter and Knowles 1978
MH-15	+	Solter and Knowles 1978
FA-25	+	Solter and Knowles 1978

<sup>1</sup> Solter and Knowles 1978

Figure 1



$\leftarrow S_1$

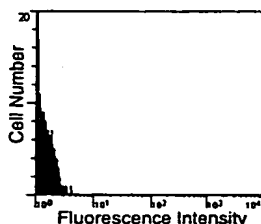
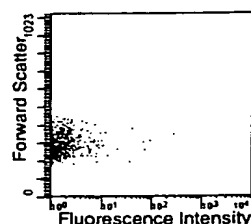
$\leftarrow S_2$

$\leftarrow S_3$

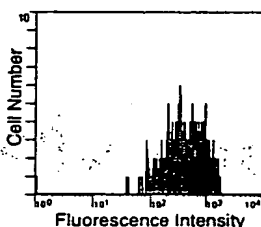
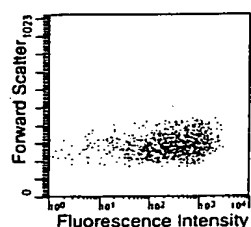
**THIS PAGE BLANK (USPTO)**

Figure 2

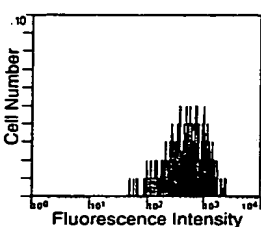
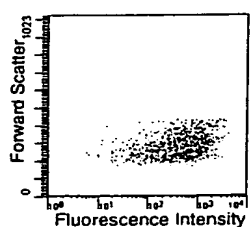
# Flow Cytofluorometric Analysis of Cell Surface Antigen Expression by 2102Ep Human EC Cells



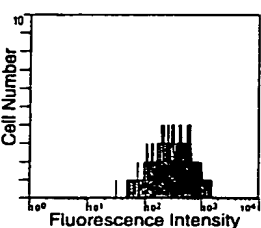
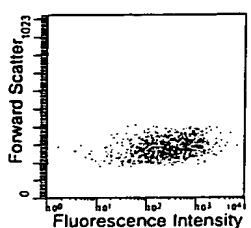
P3X (negative control)



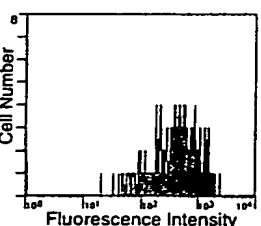
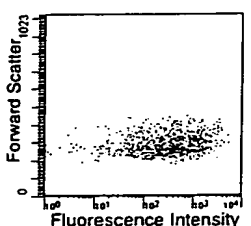
SSEA-3



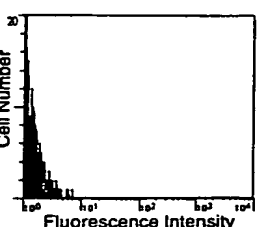
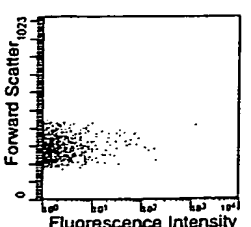
SSEA-4



TRA-1-60



TRA-1-81

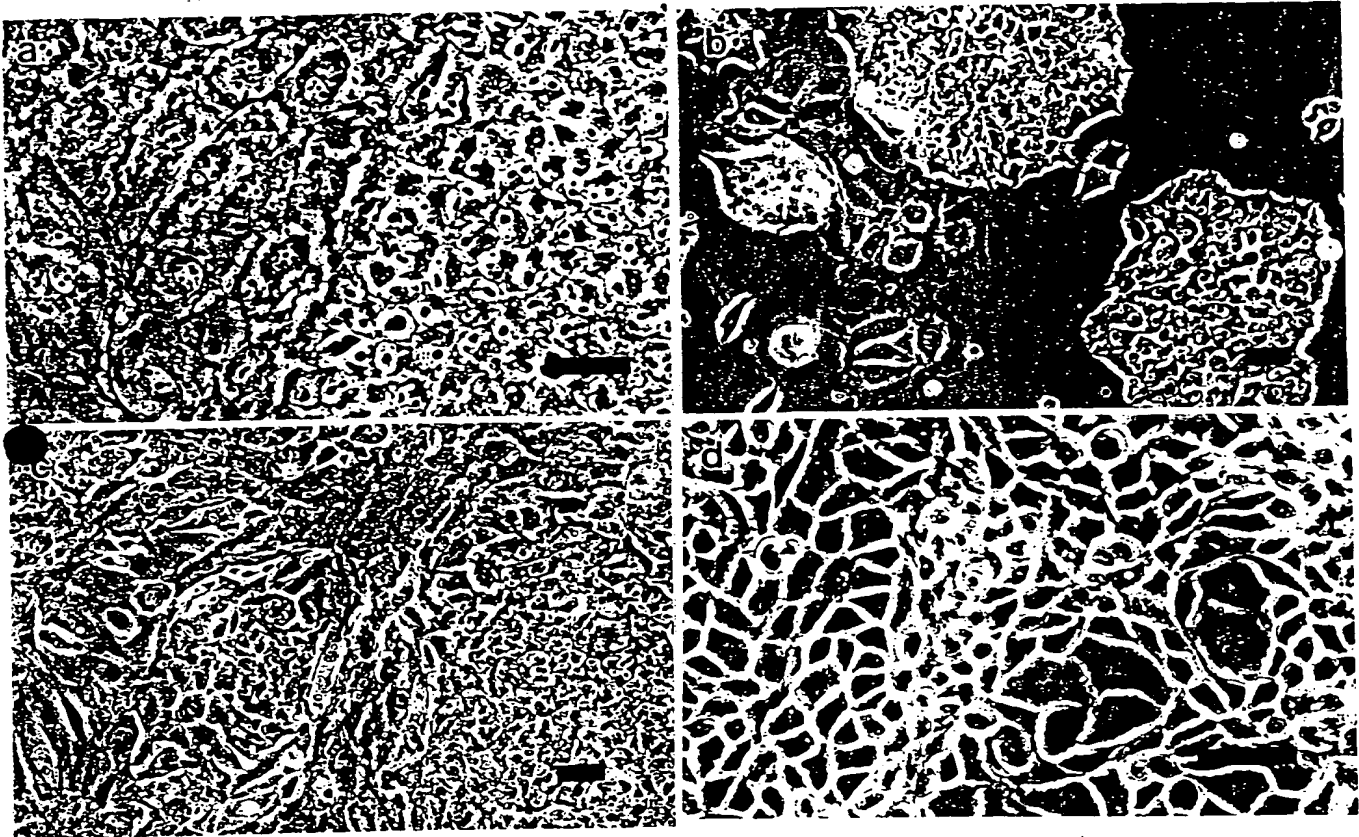


SSEA-1

**THIS PAGE BLANK (USPTO)**

Figure 3

S/O  
↓



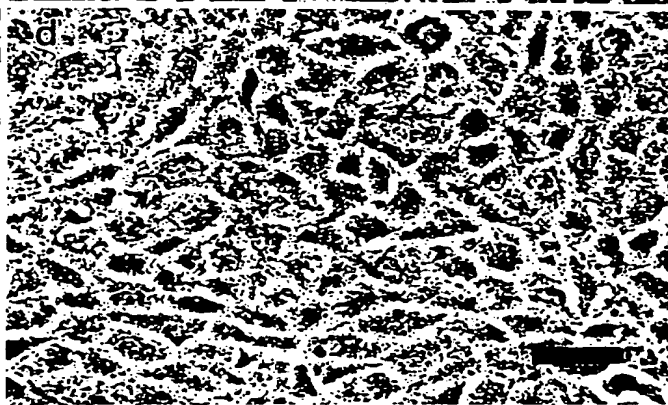
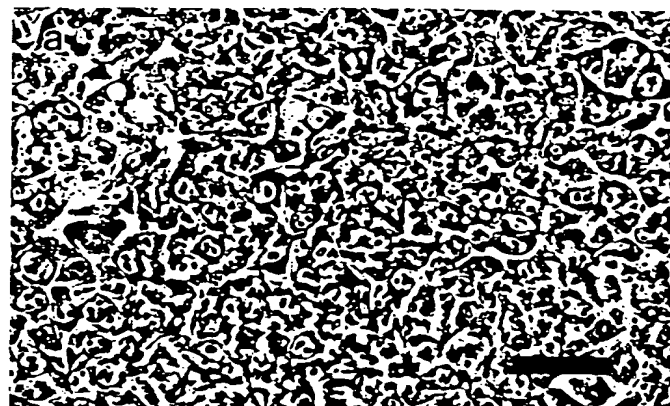
Electron Micrograph of Cells

**THIS PAGE BLANK (USPTO)**



Figure 4

s/o



← s/o

same type of tissue as in

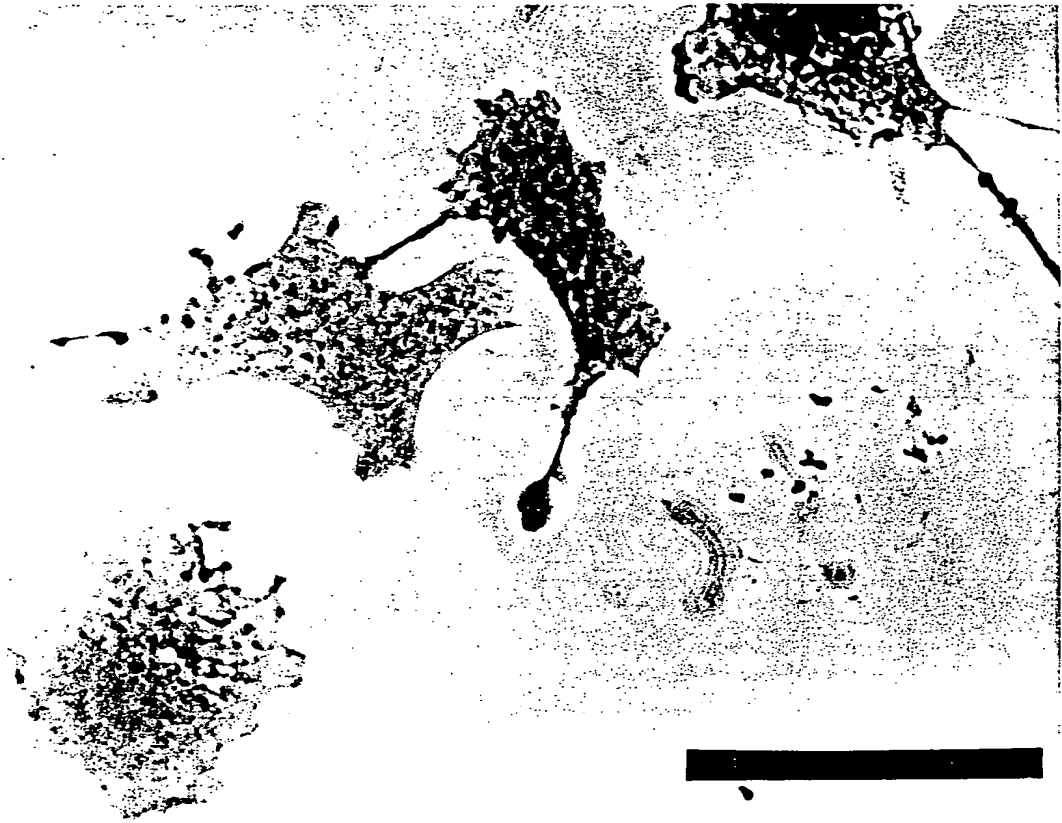
**THIS PAGE BLANK (USPTO)**

Figure 5



**THIS PAGE BLANK (USPTO)**

Figure 6



Electron Micrograph

ACT NO: GBC / 00582

Form 23/77 : 18/2/00

Agent : Harrison Goddard  
Footc.

**THIS PAGE BLANK (USPTO)**

**THIS PAGE BLANK (USPTO)**